

# Phase-Correlated Perfusion Imaging of Free-Breathing Rodents

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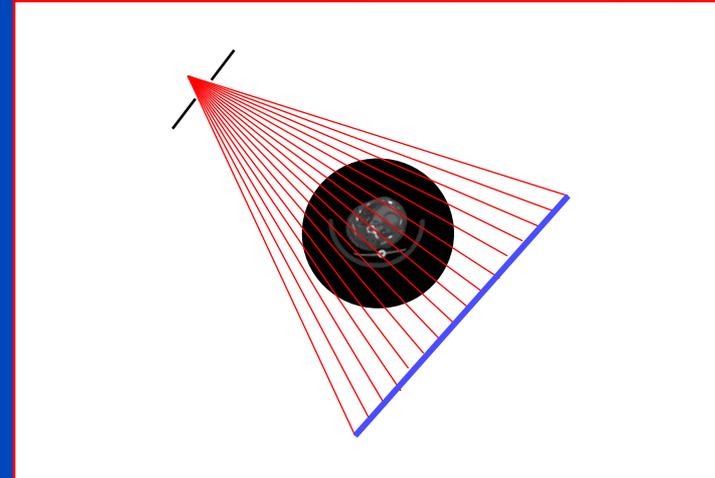
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# Aim

- Perfusion studies are already a common tool in clinical practice.
- Although many animal models for cardiac diseases exist **cardiac perfusion** is difficult in small animals due to the high cardiac and respiratory rates of up to 400 bpm and 250 rpm, respectively.
- We aim at providing a scan and acquisition protocol and dedicated reconstruction method that allows to perform perfusion studies of free breathing small animals at reasonable dose.

# VolumeCT (VCT)

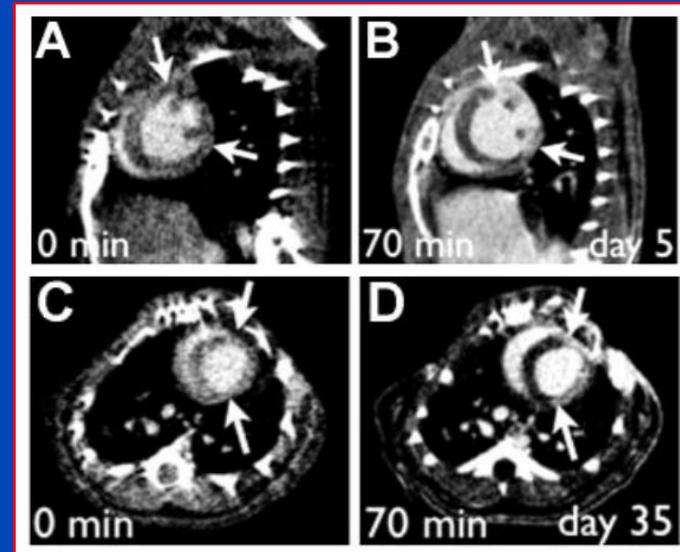
- VolumeCT (Siemens Healthcare, Forchheim, Germany)
- X-ray source:
  - Focal spot size:  $400\ \mu\text{m} \times 400\ \mu\text{m}$
  - Tube voltage range: 80 kV – 140 kV
  - Tube current range: 10 mA – 50 mA
- Detector:
  - Varian flat panel detector
  - $1024 \times 768$  pixel (2x2 binning)
  - $1024 \times 192$  @ **100 fps**
  - $388\ \mu\text{m}$  pixel size
  - Spatial sampling:  $238\ \mu\text{m}$
  - 10 ms integration time
- Protocol:
  - Scan time: 20 s
  - Rotation speed:  $18\ ^\circ/\text{s}$
  - Number of projections: 2000
  - Estimated dose: 50 mGy



# Cardiac Perfusion of Small Animals

## Prior Art

- High resolution (100  $\mu\text{m}$ ) imaging of the thoracic region.
- No phase-correlation and thus motion artifacts occur.
- Administration of more than 1 mL of contrast agent within 70 min.

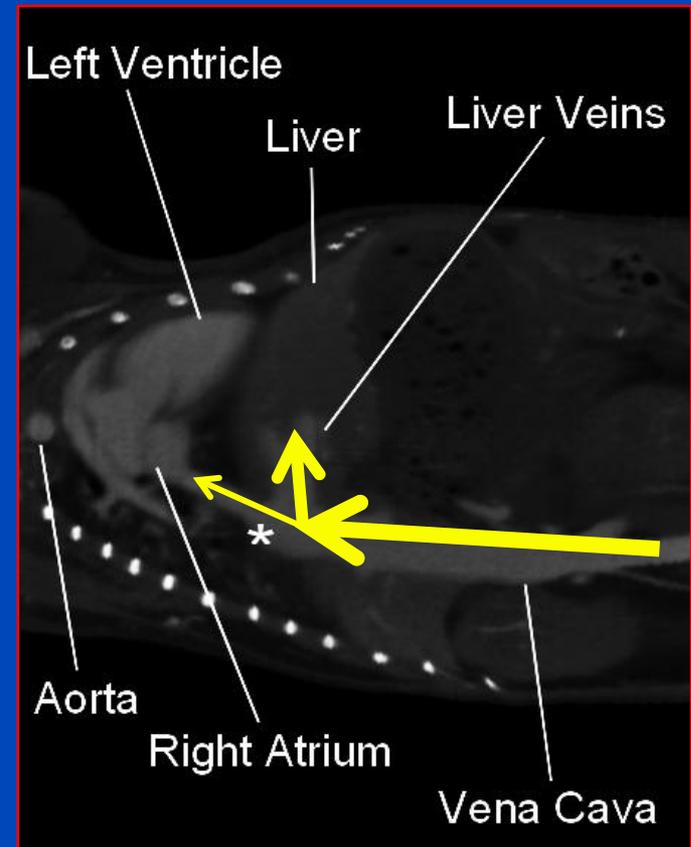


Nahrendorf M, Badea C, Hedlund LW, Figueiredo JL, Sosnovik DE, Johnson GA, Weissleder R. High-resolution imaging of murine myocardial infarction with delayed-enhancement cine micro-CT. *American Journal of Physiology: Heart and Circulatory Physiology*. 2007; 292:H3172–H3178.

# CONTRAST INJECTION PROTOCOL

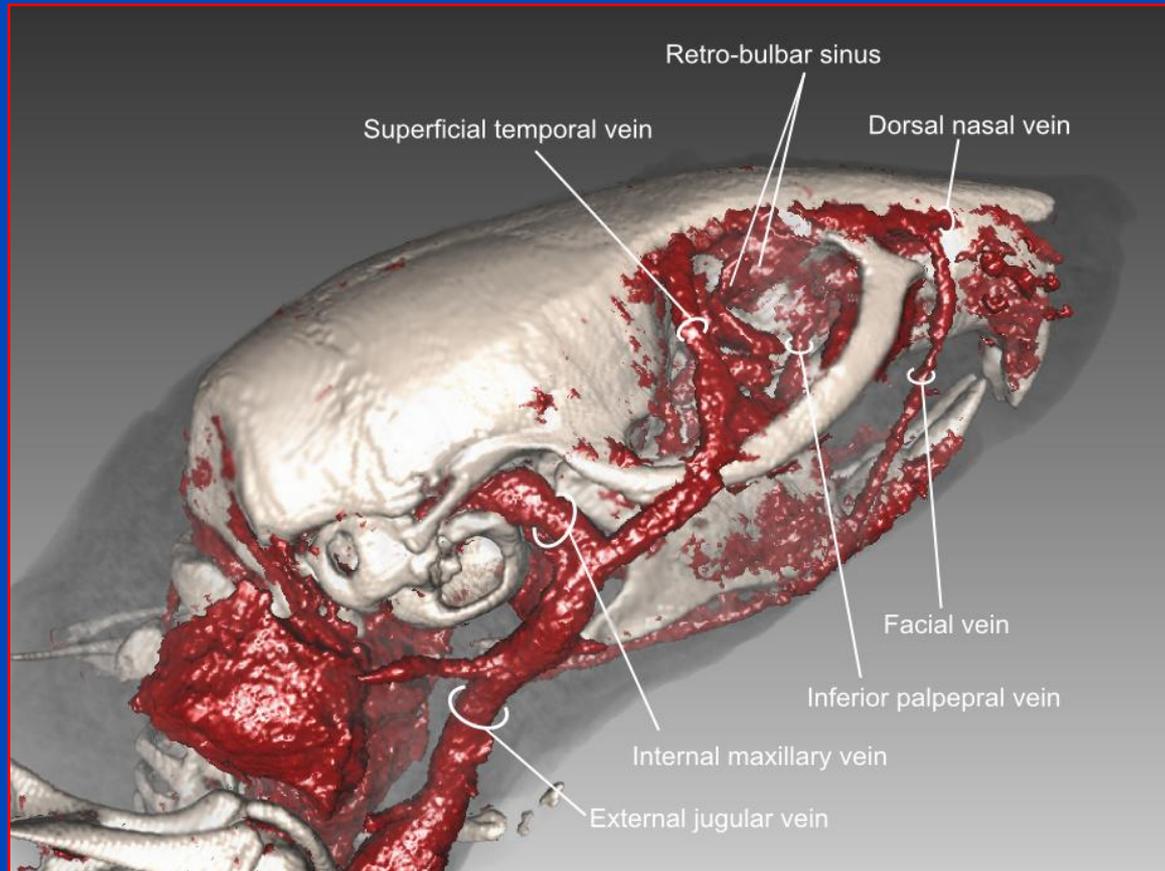
# Contrast Injection

- We wish to inject boli of 25  $\mu\text{L}$ .
- Clinical contrast agents are highly viscous (up to 8.7 mPa-s).
- Retrograd blood flow from the vena cava to the liver veins near the diaphragm.
- Bolus is dissolved before it arrives in the heart.
- Another route for contrast injection is required.
- We propose to inject into the **retro-bulbar sinus**.



Curved MPR through the vena cava of a mouse obtained from a high resolution micro-CT scan.

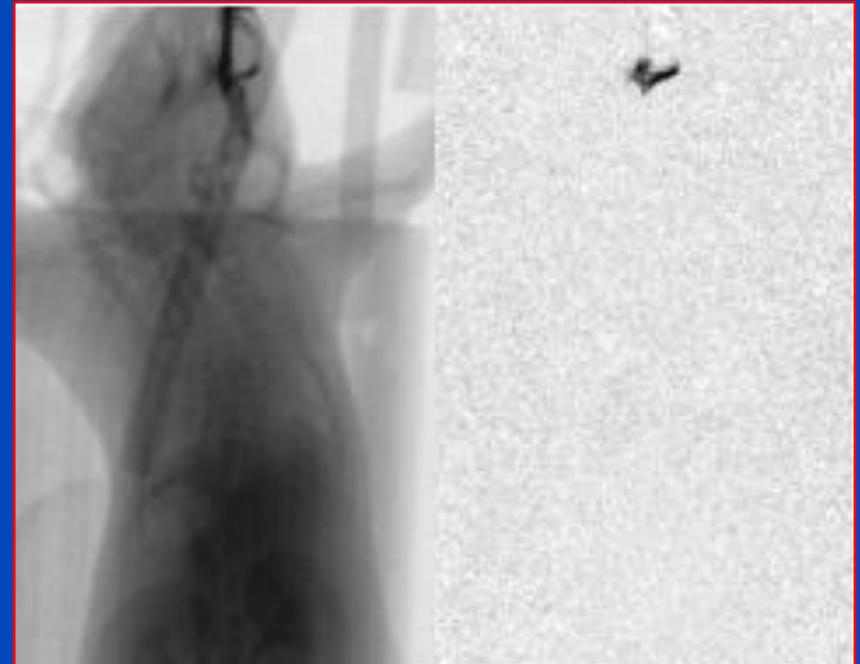
# Contrast Injection



**Volume rendering of a high resolution micro-CT scan with a spatial resolution of about 40  $\mu\text{m}$ .**

# Contrast Injection

- Injection into the retro-bulbar sinus is verified using digital subtraction angiography.
- Imeron 300 is used as contrast agent.
- Contrast agent arrives in the right ventricle 1.5 s after the injection.
- Contrast agent is in the left ventricle after about 2.0 s.
- Enhancement of the aorta visible after about 2.5 s.

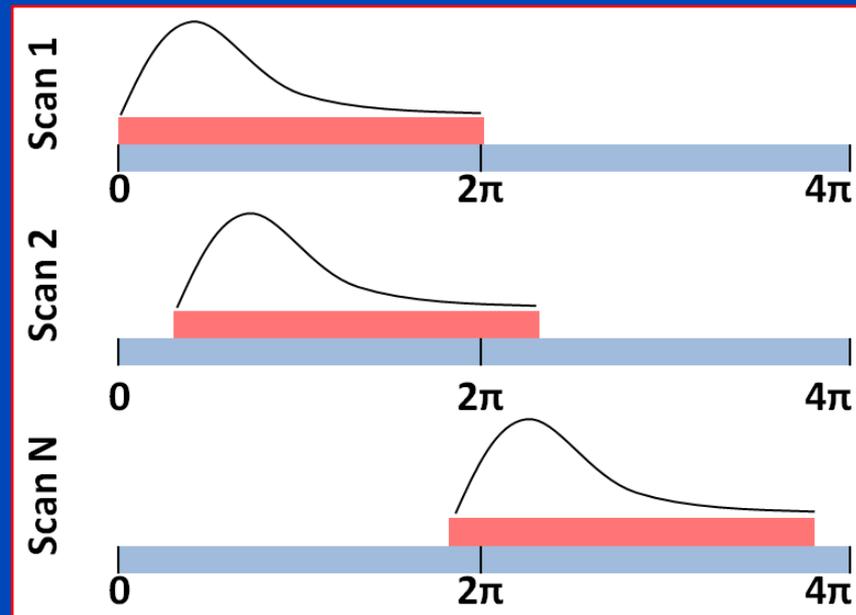


Left figure: acquired projection images.  
Right figure : subtraction angiography.

# SCAN PROTOCOL & GATING

# Scan Protocol

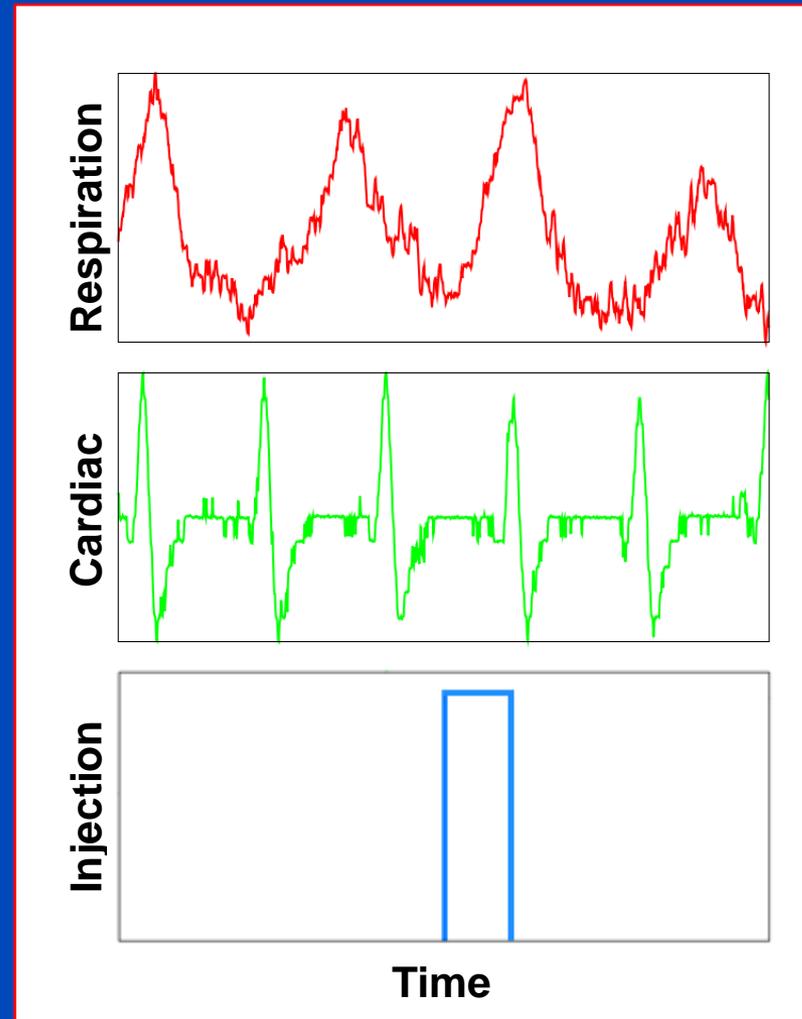
- We perform  $N=10$  scans each over  $360^\circ$  within 20 s.
- 2000 projections are acquired in every scan.
- Each scan starts at a different angle. We thus ensure to cover the complete angular range.
- We inject  $25\ \mu\text{L}$  per scan and  $250\ \mu\text{L}$  in total.



Schematic illustration of the used scan protocol. This is inspired by *Badea CT, Johnston SM, Subashi E, Qi Y, Hedlund LW, Johnson GA. Lung perfusion imaging in small animals using 4D micro-CT at heartbeat temporal resolution. Medical Physics. 2010; 37:54–62.*

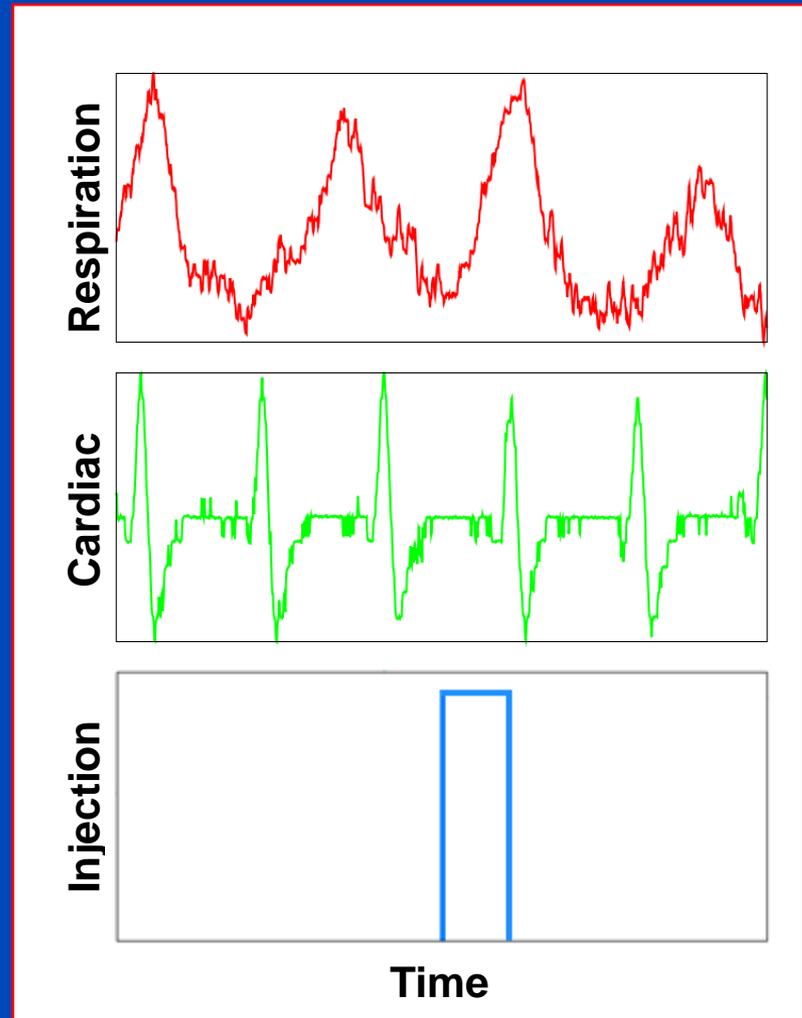
# Extrinsic Gating

- Respiration (r) is monitored using a pneumatic pillow.
- Information on cardiac motion (c) are obtained using electrodes attached to the paws.
- Timestamp of contrast injection/start of the perfusion (p) is recorded.
- All signals are retrospectively correlated to the acquired projections.
- Phase windows for image reconstruction are defined by c, r, p and corresponding window widths  $\Delta c$ ,  $\Delta r$ ,  $\Delta p$ .



# Extrinsic Gating

- Three gating signals require the reconstruction to be correlated to all of these signals.
- Example:
  - 10×2000 projections in total
  - Cardiac window: 20 %
  - Respiratory window: 20 %
  - Perfusion window: 1 s (5 %)
  - Only 0.2 % of all acquired projections contribute to a reconstruction.



# IMAGE RECONSTRUCTION

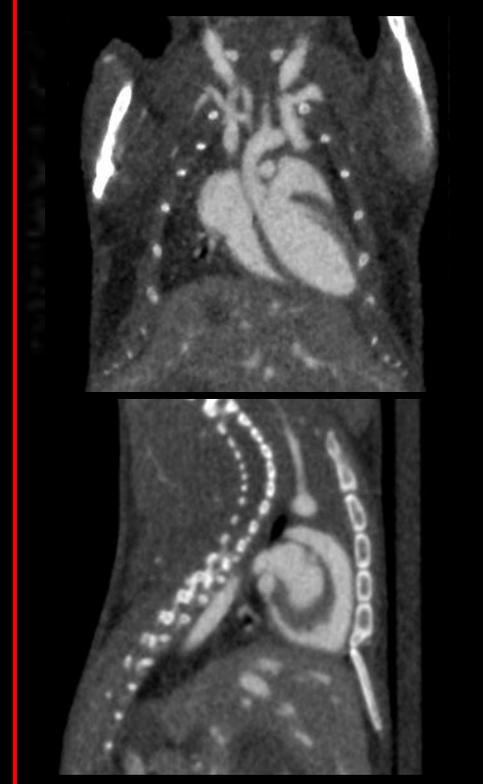
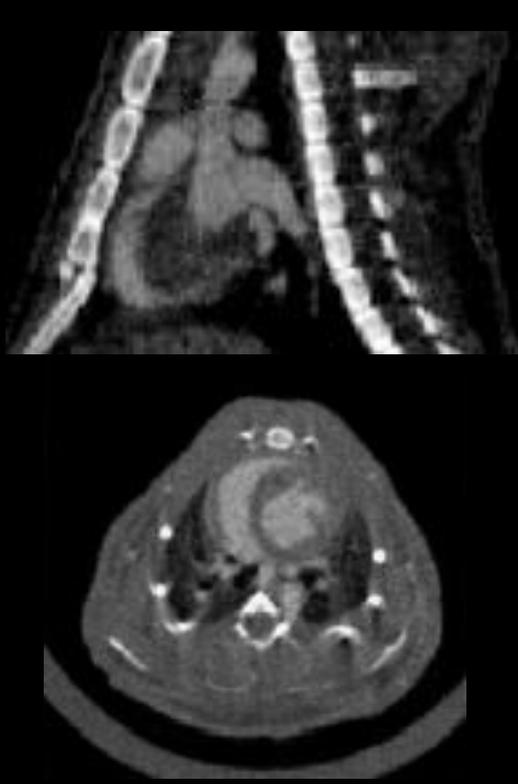
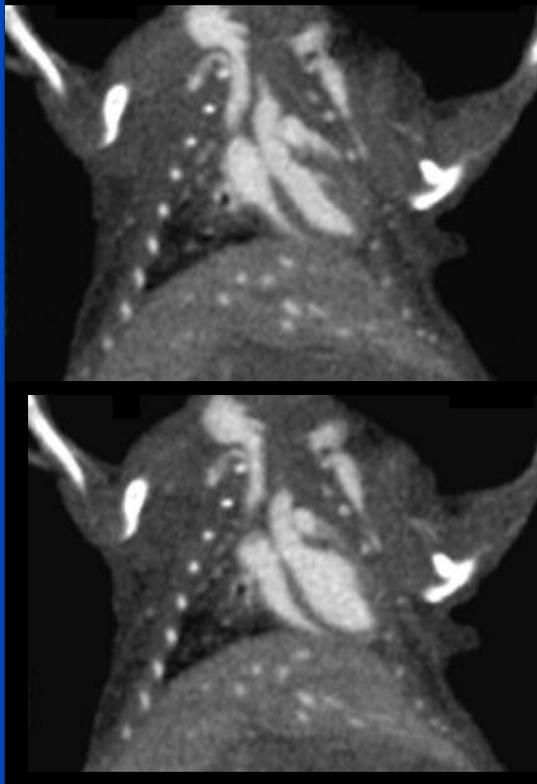
## THE PLDPC ALGORITHM

# Image Reconstruction Prior Art

280 mGy, 150  $\mu$ m, 10 phases

1840 mGy, 90  $\mu$ m, 12 phases

500 mGy, 80  $\mu$ m, 50 phases

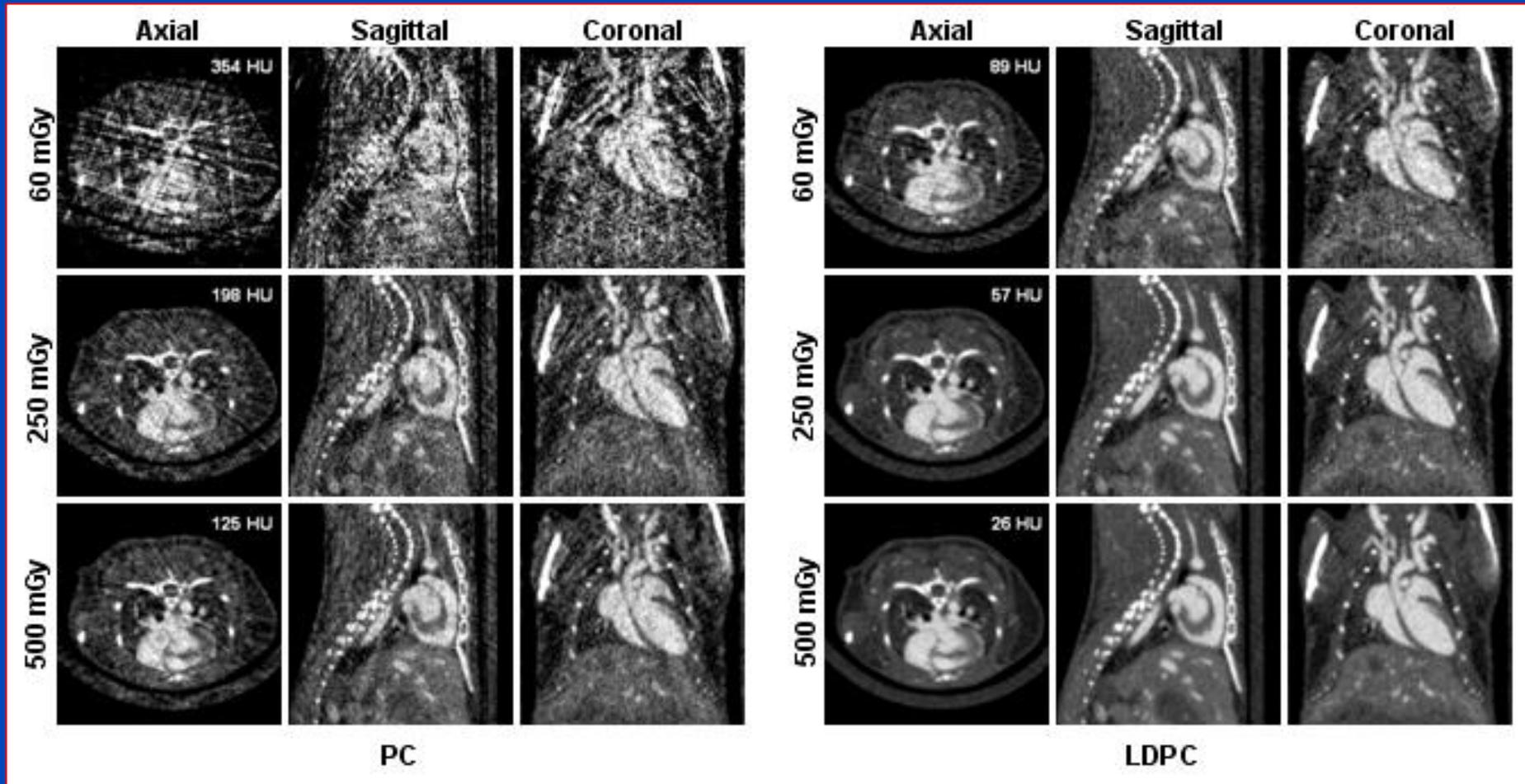


M. Drangova, N. L. Ford, S. A. Detombe, A. R. Wheatley, and D. W. Holdsworth, "Fast retrospectively gated quantitative four-dimensional (4D) cardiac micro computed tomography imaging of free-breathing mice," *Investigative Radiology*, vol. 42, no. 2, pp. 85–94, Feb. 2007.

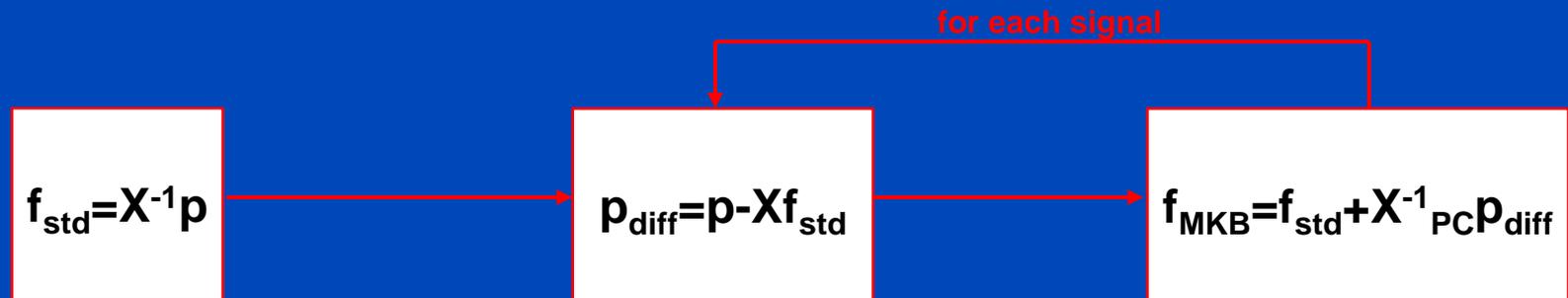
C. Badea, B. Fubara, L. Hedlund, and G. Johnson, "4D micro-CT of the mouse heart," *Molecular Imaging*, vol. 4, no. 2, pp. 110–116, Apr./Jun. 2005.

S. Sawall, F. Bergner, R. Lapp, M. Mronz, M. Karolczak, A. Hess, and M. Kachelrieß, "Low-dose cardio-respiratory phase-correlated cone-beam micro-CT of small animals," *Medical Physics*, vol. 38, no. 3, pp. 1416–1424, Feb. 2011.

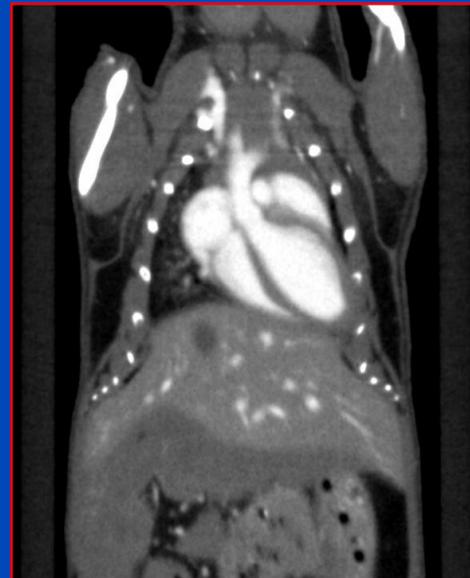
# Image Reconstruction Prior Art



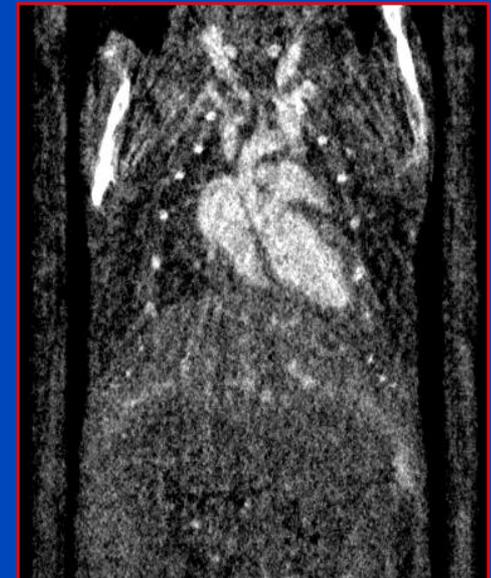
# Modified McKinnon-Bates Algorithm



- Use image based on all projections as prior (standard image)
- Calculate rawdata difference for desired motion phases
- Perform correction



Standard image  $f_{\text{std}}$  reconstructed from all projections.

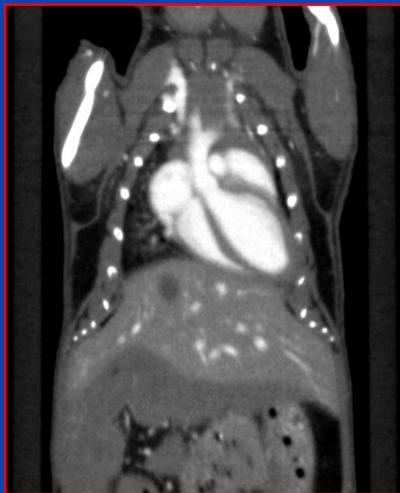


McKinnon-Bates reconstruction  $f_{\text{MKB}}$ .

# Edge Preserving Spatio-Temporal Postprocessing

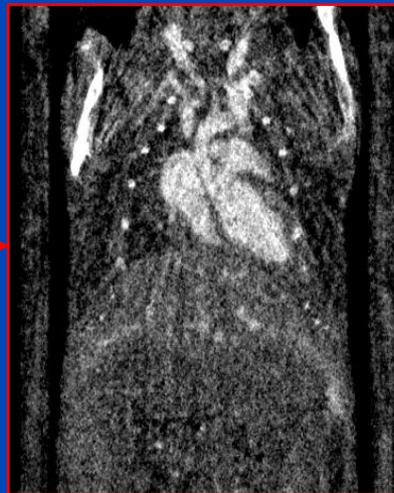
Five-dimensional bilateral filtering (three spatial and two temporal dimensions)

$$f_{\text{LDPC}} = Bf_{\text{MKB}} = \frac{\int dt^5 D(x,t)R(x,t)f(t)}{\int dt^5 D(x,t)R(x,t)}$$



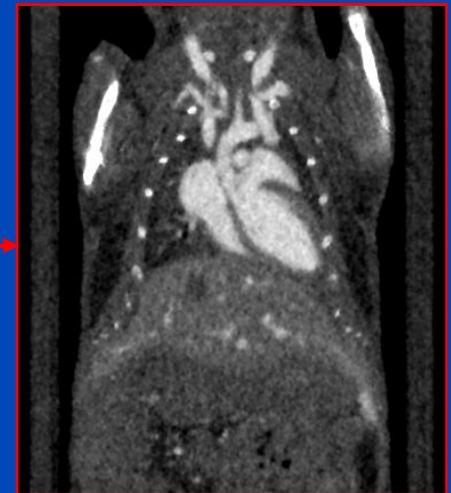
Standard image reconstructed from all projections.

Modified  
MKB



Modified McKinnon-Bates reconstruction.

5D Filter

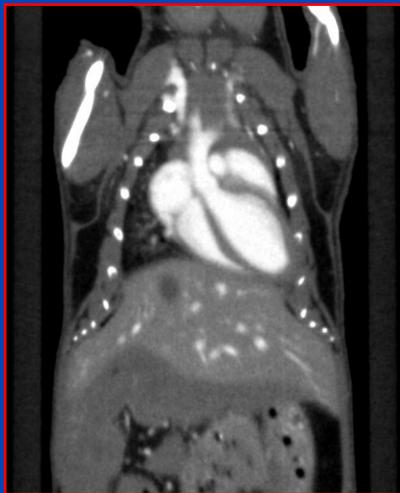


Final low-dose phase-correlated (LDPC) image.

# Edge Preserving Spatio-Temporal Postprocessing

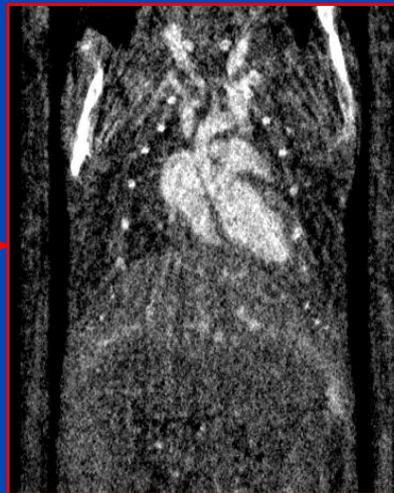
Six-dimensional bilateral filtering (three spatial and three temporal dimensions)

$$f_{\text{PLDPC}} = B f_{\text{MKB}} = \frac{\int dt^6 D(x,t) R(x,t) f(t)}{\int dt^6 D(x,t) R(x,t)}$$



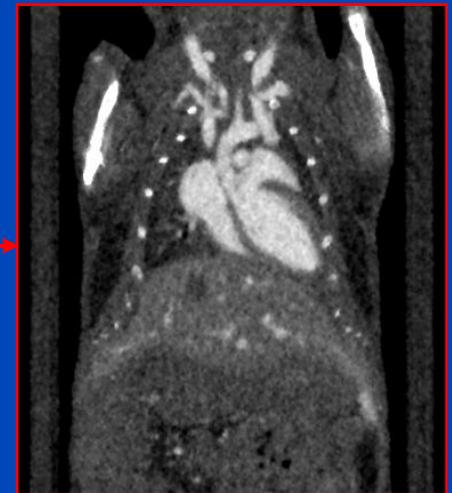
Standard image reconstructed from all projections.

Modified MKB



Modified McKinnon-Bates reconstruction.

6D Filter



Final low-dose phase-correlated (PLDPC) image.

# RESULTS

# Results

	Mouse 1	Mouse 2
Respiratory rate	120 rpm	115 rpm
Cardiac rate	265 bpm	250 bpm
Contrast agent	Imeron 300	Imeron 300
Administered volume	10x25 µL	10x25 µL

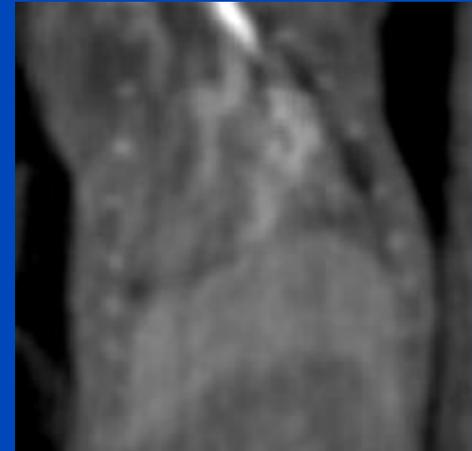
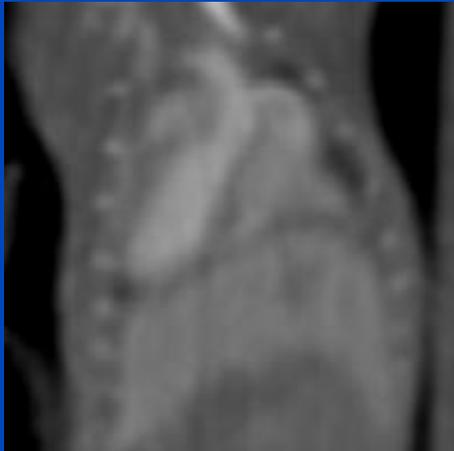
# Mouse 1

Respiration

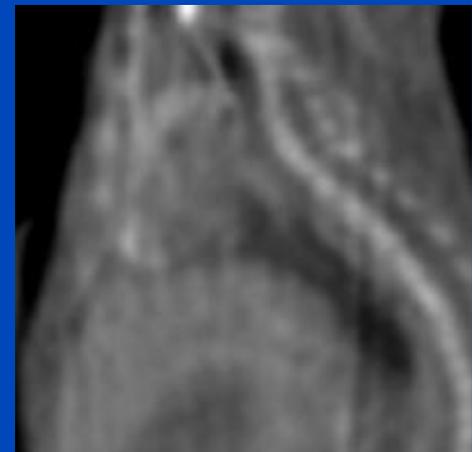
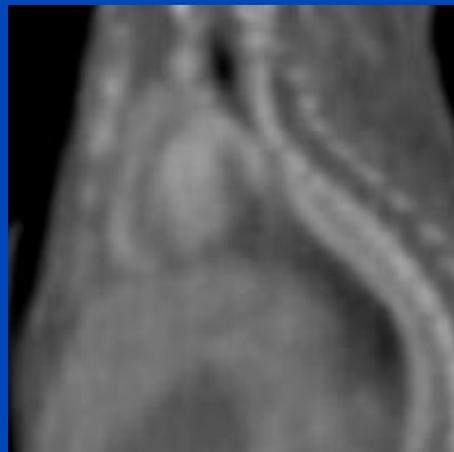
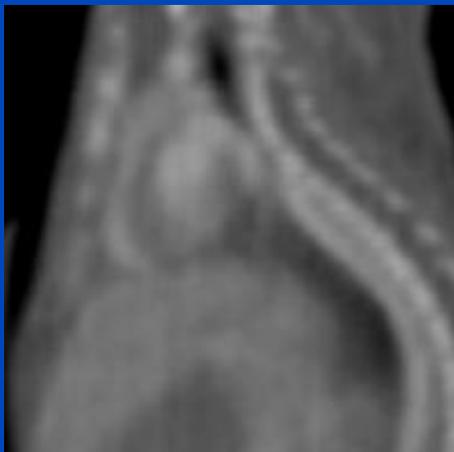
Cardiac Motion

Perfusion

Coronal



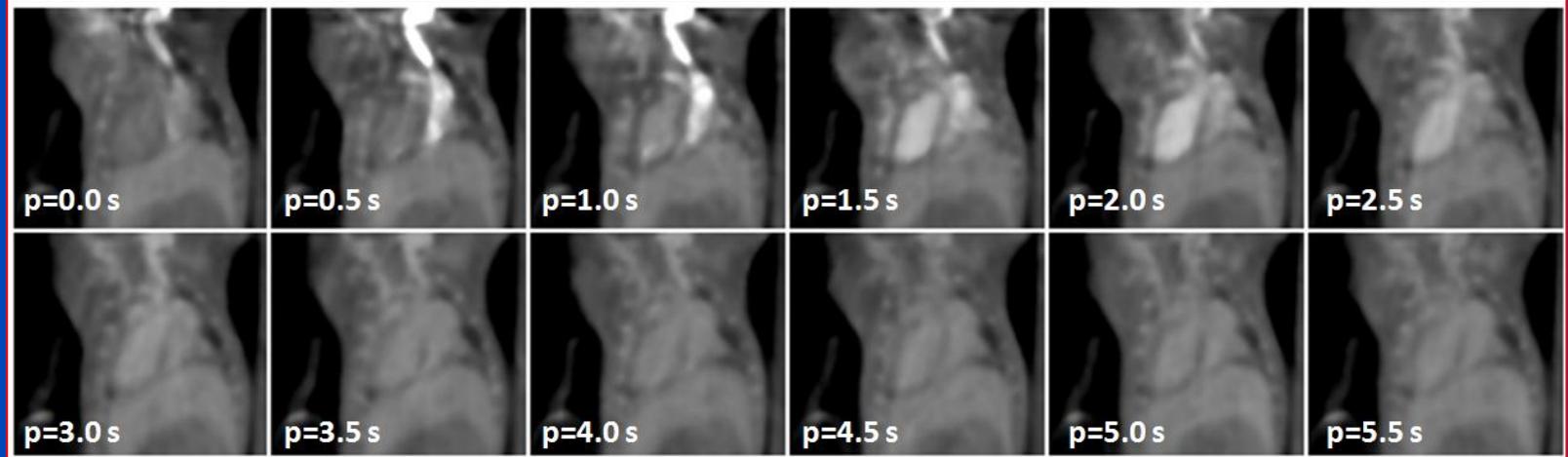
Sagittal



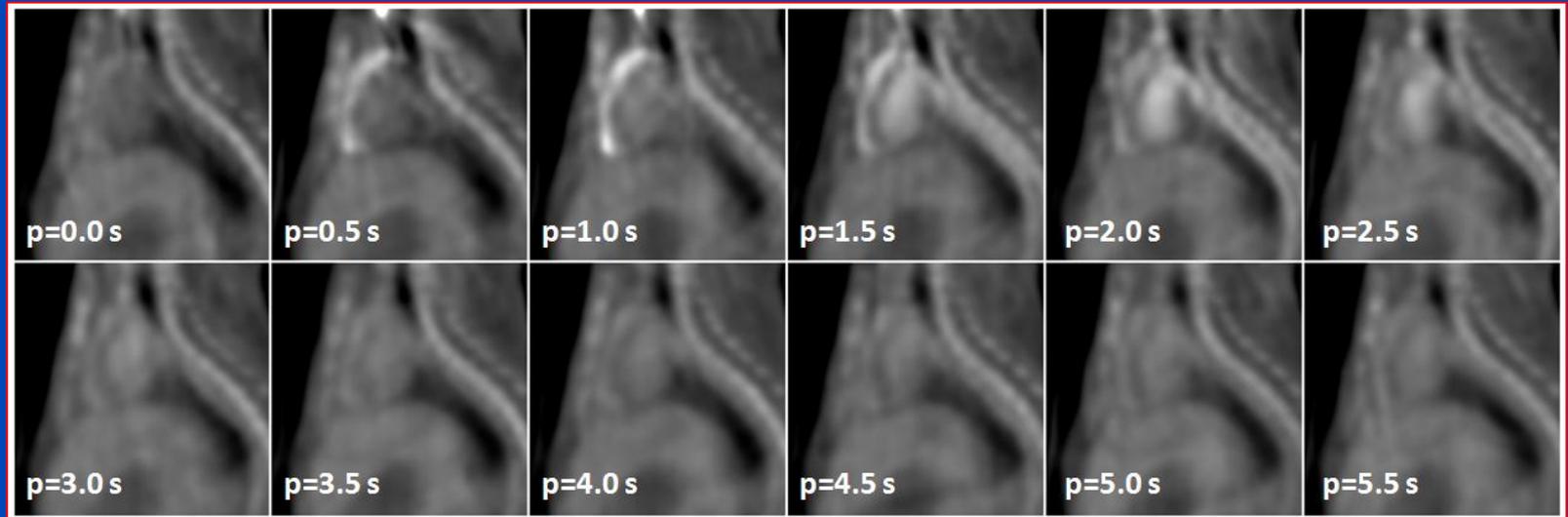
$\Delta r=20$ ,  $\Delta c=20\%$ ,  $\Delta p=0.5$  s (600 HU / 700 HU)

# Mouse 2

Coronal

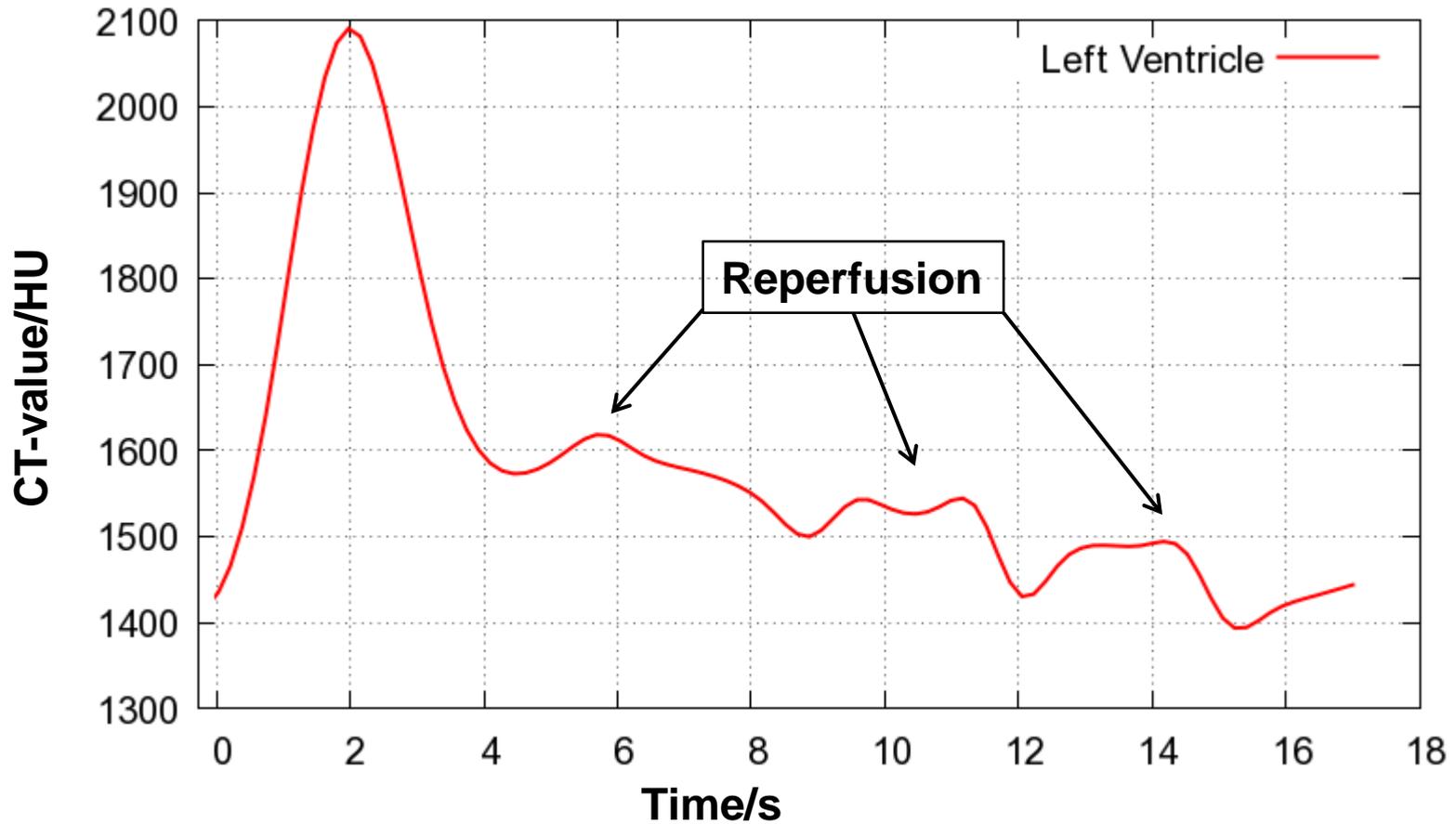


Sagittal

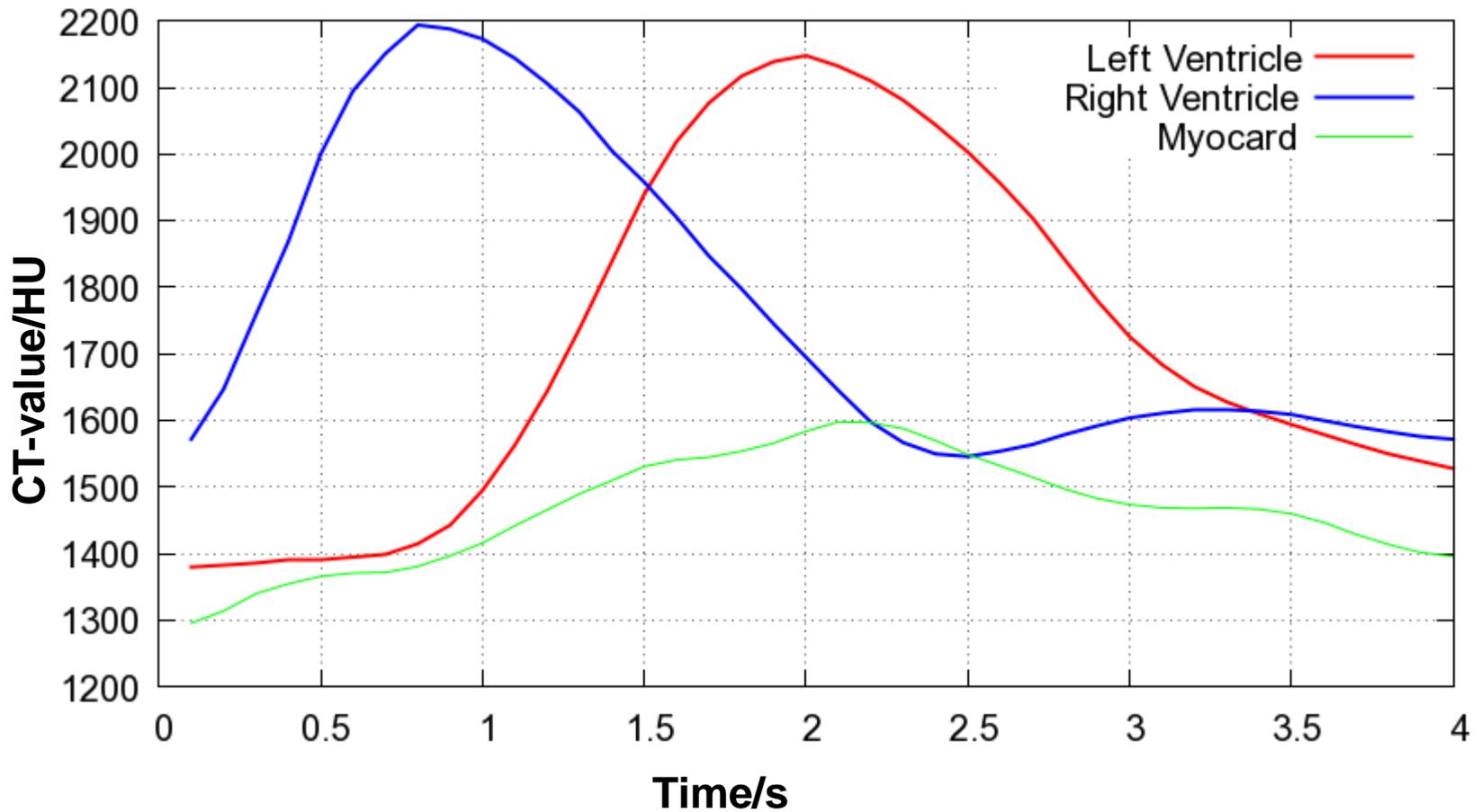


$r=0\%$ ,  $\Delta r=20\%$ ,  $c=20\%$ ,  $\Delta c=20\%$ ,  $\Delta p=0.5$  s (600 HU / 900 HU)

# Time-Density-Curve



# Time-Density-Curve



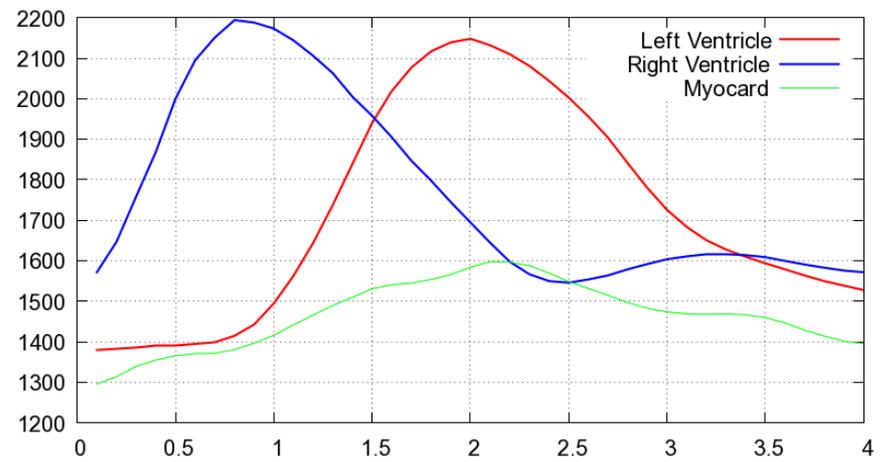
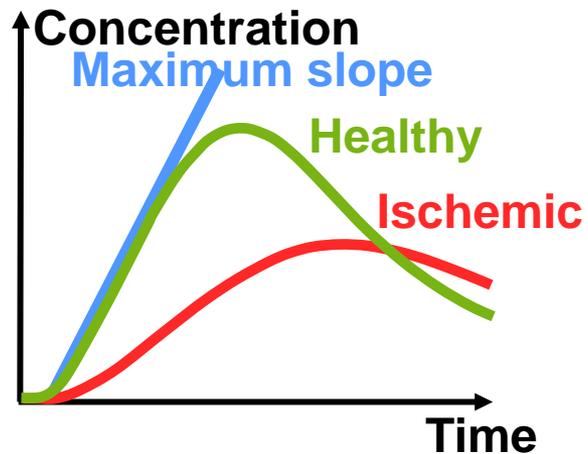
# Clinical Case

## Clinical Examinations

SOMATOM Definition Flash  
Flash Speed. Lowest Dose.



## Preclinical Examinations



# Summary & Conclusions

- Cardiac- and respiratory-correlated reconstructions come at no additional cost allowing for the quantification of ejection fraction etc.
- The dose per imaging study is about 500 mGy, what is far below the LD<sub>50</sub> of 5-7 Gy.
- The injection technique is minimally invasive allowing for longitudinal studies.
- The quantitative results correspond well to what is known from clinical practice.
- Overall results show that cardiac perfusion studies in small rodents are possible.
- This boosts preclinical research with a lot of new possibilities.

# Thank you!

This presentation will shortly be available at [www.dkfz.de/ct](http://www.dkfz.de/ct).

This work was supported by the **Deutsche Forschungsgemeinschaft** (DFG) under grant FOR 661. The high-speed image reconstruction software RayConStruct-IR was provided by **RayConStruct®** GmbH, Nürnberg, Germany. The authors thank the **Intel** Corporation and **Fujitsu** Technology Solutions GmbH for providing their latest multicore hardware. The authors further thank **Sandra Strobel**, **Johannes Käßer** and **PD Dr. Andreas Hess** for help with the mouse measurements. The blood pool contrast agent ExiTron nano 12000 was provided by **Miltenyi Biotec** GmbH, Bergisch Gladbach, Germany and **NanoPET** GmbH, Berlin, Germany. The high-precision pump was assembled by **Klaus Schewiola** and **Steffen Seeber** at the DKFZ.