

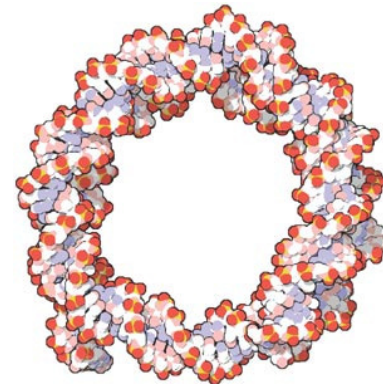
# **High flexibility of DNA on short length scales probed by atomic force microscopy**

**Wiggins P. A. et al.  
Nature Nanotechnology (2006)**

**presented by Anja Schwäger  
23.01.2008**

# Outline

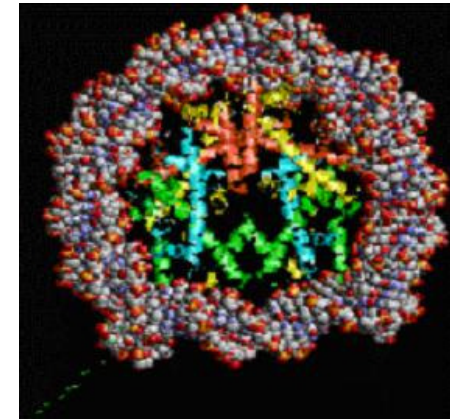
- Theory/Background
  - Elasticity of DNA
  - Aim of study
- Methods
  - AFM
  - Data analysis
- Results
  - New model for DNA elasticity
  - Comparison with other results
- Outlook
  - Improvement of model
  - Summary



# DNA mechanics

## DNA elasticity

- Bending and twisting properties of DNA
  - ⇒ determine DNA conformation and dynamics
- Elastic properties of DNA
  - double helix relatively stiff for bending
  - unlikely to form sharp bends, high energy needed
- Biological importance of sharp DNA bends on short length scales
  - chromatin
  - packaging of DNA in capsid
  - protein interactions (gene regulation)



Nucleosome

<http://www.jyi.org/research/re.php?id=419>

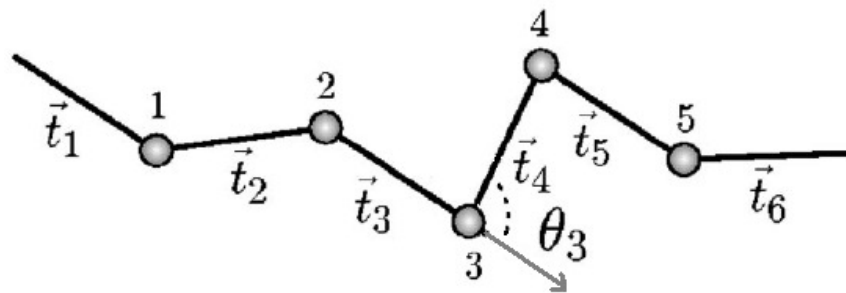


Lac Operator

Swigon, PNAS, 2006

# Worm-Like Chain (WLC)

- **Elasticity model** for the description of **semiflexible polymers**
- Molecule as a thin rod with constant and isotrop elastic behaviour
- Approximation (discrete segments):
  - Conformation described as a chain of segments
  - Each segment with classical elastic energy cost for bending (Hooke's law)
- WLC is this approximation in the limit of zero segment length



modified according to Wiggins and Nelson, Phys Rev E, 2005

# Worm-Like Chain (WLC)

- Important parameter of the WLC:

**Persistence length** of a polymer

Measure for molecule stiffness / the resistance of the chain to bending

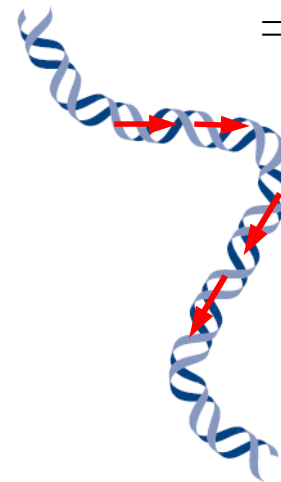
(length over which correlation of tangent vector directions is lost  
 $\approx$  distance that separates bends)

**short** persistence length  
 $\Rightarrow$  more flexible



**long** persistence length

$\Rightarrow$  stiffer



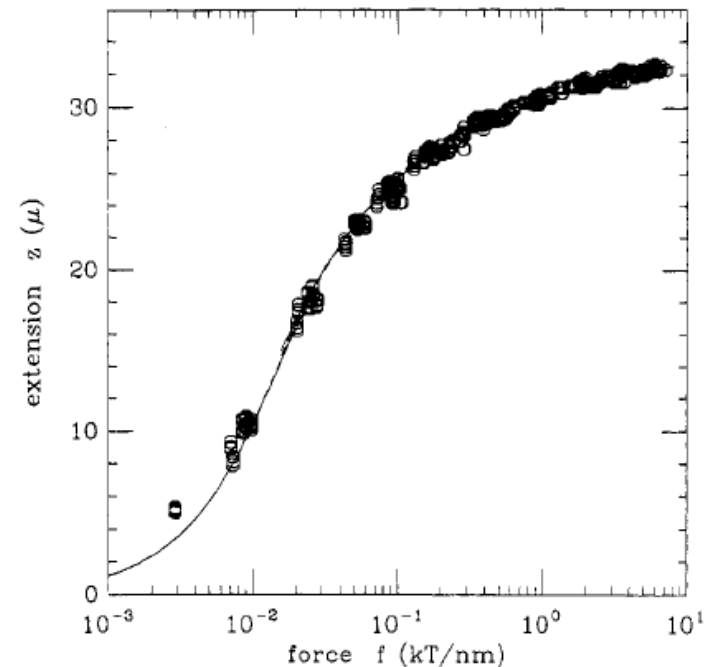
Podgornik, Nature Nanotechnology, 2006

# WLC description of DNA

## Example: Stretching of DNA



- Simple description of the behaviour of a polymer chain as response to extension force
  - reduction of entropy with straightening
- Length scale longer than persistence length
  - results for  $L = 32.8 \mu\text{m}$
  - persistence length of double helix DNA  $L_p \approx 50 \text{ nm (150 bp)}$



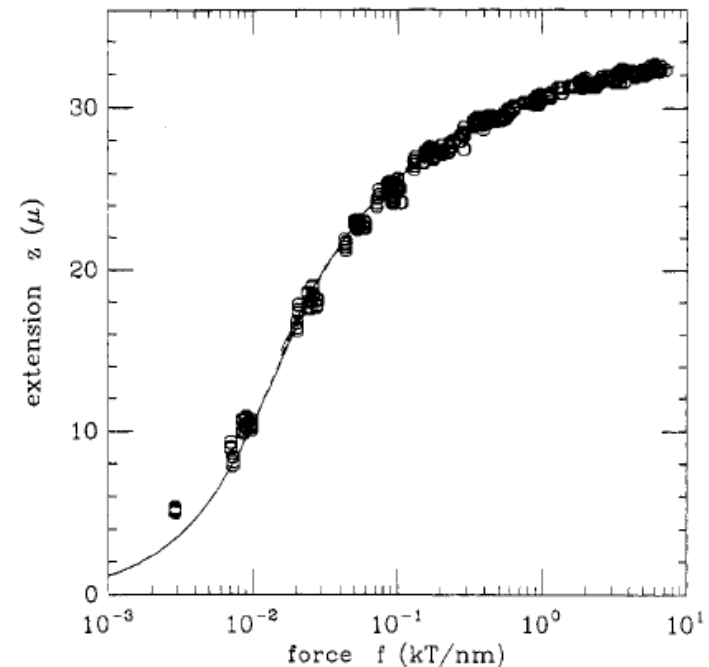
Marko and Siggia, Macromolecules, 1995

# WLC description of DNA

## Example: Stretching of DNA



- Simple description of the behaviour of a polymer chain as response to extension force
  - reduction of entropy with straightening
- Length scale longer than persistence length
  - results for  $L = 32.8 \mu\text{m}$
  - persistence length of double helix DNA  $L_p \approx 50 \text{ nm (150 bp)}$



Marko and Siggia, Macromolecules, 1995

# WLC description of DNA

## Bending energy of a conformation

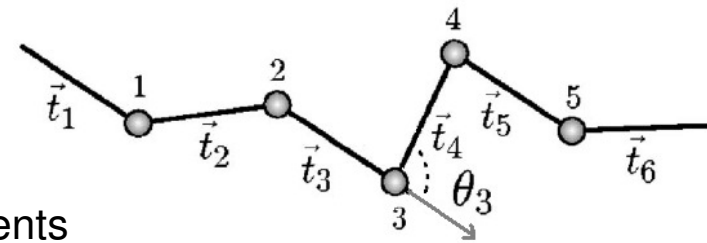
Chain of segments of length  $\ell$  with **harmonic energy** cost per segment:

$$E_{\text{WLC}}(\theta) = \frac{1}{2} k_B T (L_P/\ell) (\theta_i)^2$$

$\theta$ : bending angle between segments

$L_P$ : persistence length of DNA

$k_B T$ : thermal energy



modified according to Wiggins and Nelson, Phys Rev E, 2005

**Energy rises quadratically with bending angle**

⇒ **Boltzmann distribution**: large bending angles will occur rarely

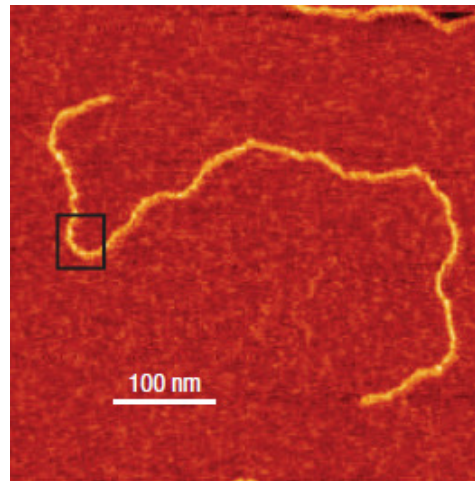


# Motivation of experiments

- Previous studies of DNA mechanics focus on longer length scales
  - Cyclization studies of **short DNA fragments**: short circles form much more easily than expected from WLC theory
  - **Test WLC** model on short length scales (few nanometers)
- ⇒ Measure model predicted **bending angle frequencies** directly in equilibrium DNA conformations
- ⇒ Frequencies of bending angles correspond to **bending energy function**

# Experimental approach

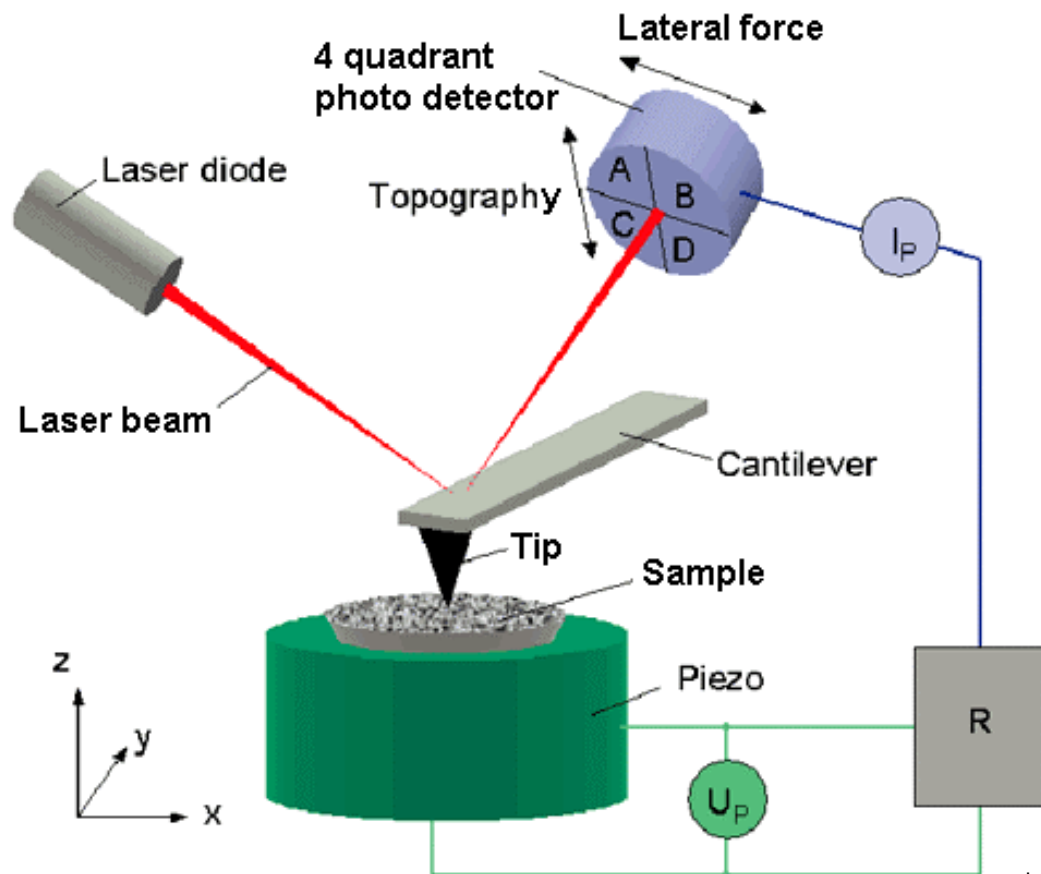
- High-resolution **atomic force microscopy**
- 2743 bp **linear double-stranded DNA** with random sequence, **adsorbed to surface**
- **Two-dimensional** movement possible
- Assume **equilibrium conformation** of adsorbed molecules



Wiggins, Nature Nanotechnology, 2006

# Atomic Force Microscopy (AFM)

## Principle



## Characteristics

- topographical images
- near angstrom resolution (0.1 nm)

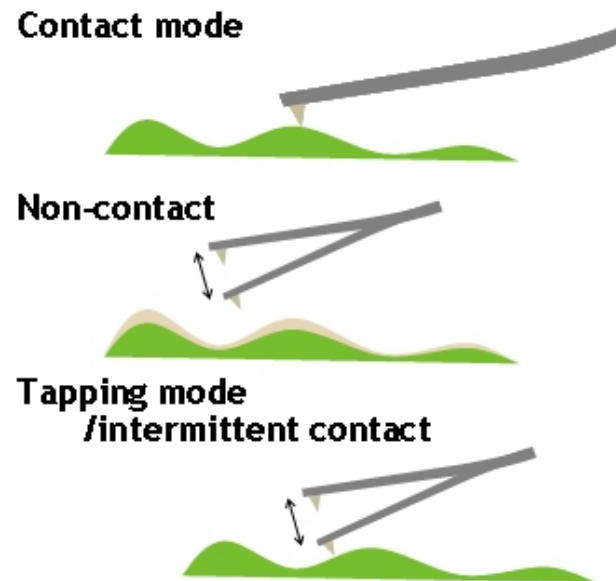
## Biological application

- imaging of living cells
- measurement in physiological buffers

<http://www.nanomat.de/datenblaetter/42000.htm>

# AFM operation modes

## Typical modes



<http://en.wikibooks.org/wiki/Nanotechnology/AFM>

- Tapping mode usually used for soft biological materials

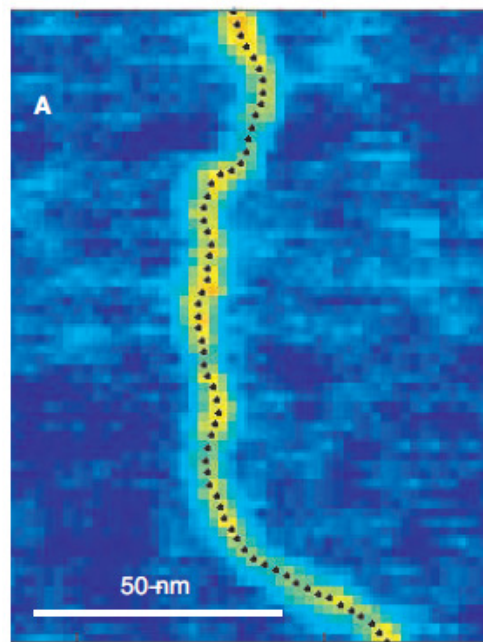
# Experimental and simulated AFM data

- Linear DNA molecules adsorbed to mica (silicate surface) with different concentrations of  $Mg^{2+}$  ions
  - AFM measurement in tapping mode **in air**
    - height profile of equilibrium conformations
  - Simulation of AFM data on the basis of WLC
    - generation of WLC conformations, 2D chains with bends
    - bending angle distribution according to WLC prediction
    - conversion to simulated AFM images
    - same analysis as experimental conformations
- ⇒ Exclude that conformations are artifacts of AFM measurement

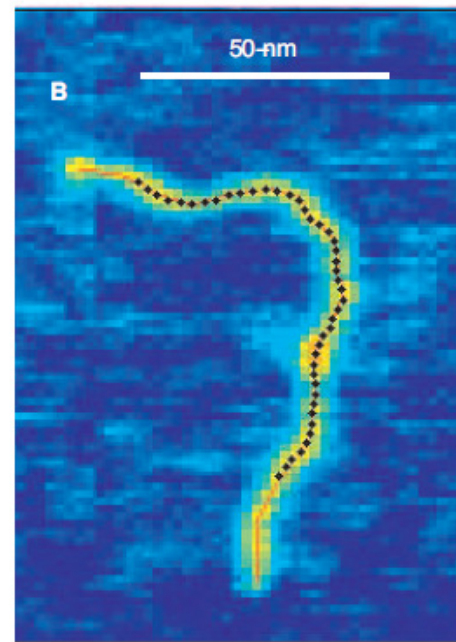
# Image analysis

## Automatic tracing of DNA contours

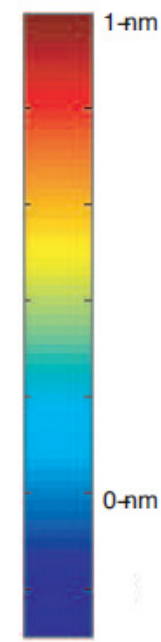
- Set points with a distance of 2.5 nm on the contour
- Length  $\ell$  of segments 2.5 nm



experimental image



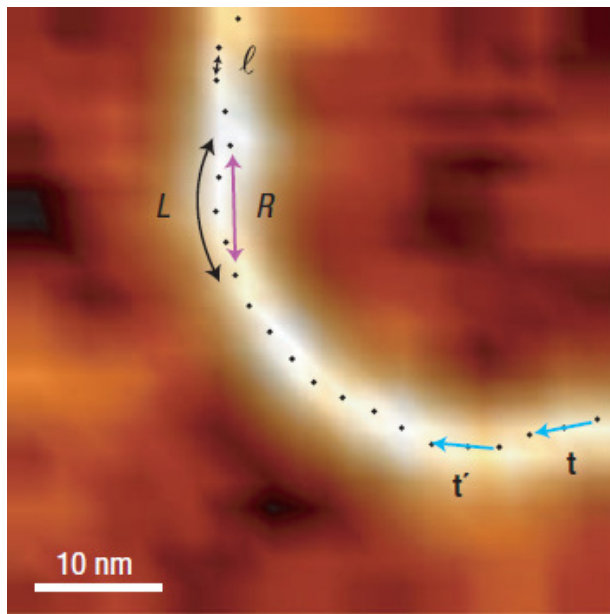
simulated image



Wiggins, Nature Nanotechnology, 2006

# Measurement of angles

- Two points of chain separated by the **contour length  $L$**
- **Tangents** of these points  $t$  and  $t'$
- Angle between two tangents
- Measured for all points on complete chain and different  $L$



Example:  $L = 3 \times 2.5$  nm  
contour length between  $t$  and  $t'$

Wiggins, Nature Nanotechnology, 2006

# Statistical evaluation

- **Angle Distribution  $G(\theta; L)$ :**  
Probability distribution of angles between tangents
  - angles between tangents  $t_i$  and  $t_{i+1}$  separated by  $L$  measured for many molecules
  - histogram of angles
- Curve measures the **bending energy  $E(\theta)/k_B T$**  ( $E(\theta)$  in units of  $k_B T$ ) of the segments of length  $L$

Bending angle distribution:

$$g(t_{i+1}|t_i) = q^{-1} \exp[-E(\theta_i)/k_B T]$$

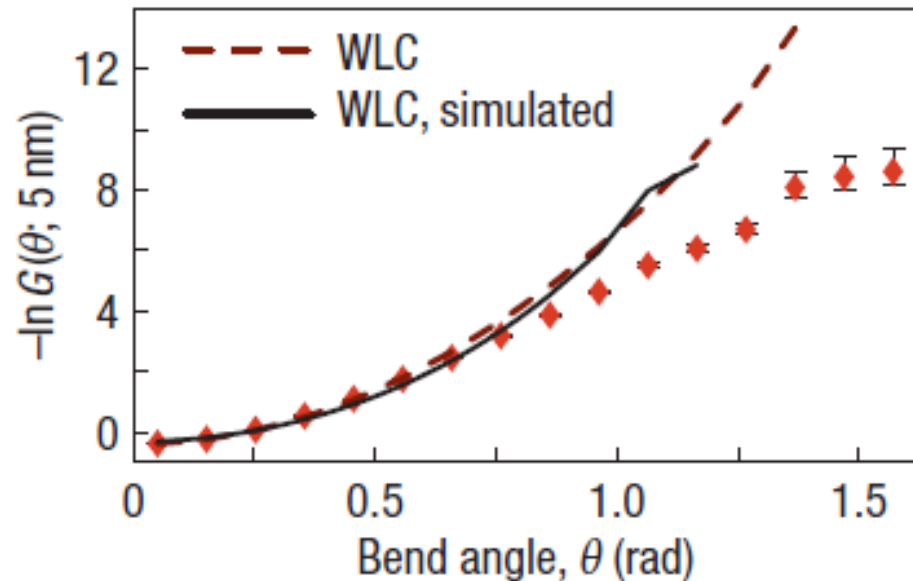
Negative logarithm:

$$-\ln(g) = \ln(q) + E(\theta_i)/k_B T$$



# Results

**Dots:** measured angle distribution for  $L = 5$  nm (negative logarithm)



Wiggins, Nature Nanotechnology, 2006

- Distribution calculated from theoretical model (WLC) does not match the measured data for large angles
- Values of measured data are much lower (= higher frequency of large angles)
- Simulated WLC data match theoretical curve

# Results

- Segments with large bends much more frequent than predicted by WLC
    - large bends ( $\geq 1.1$  rad) 30 times more frequent
  - Elastic energy for high bending angles must be lower than predicted
  - Energy function is almost linear
- ⇒ Hooke's law does not apply for the elastic energy on small length scale

# Other experiments

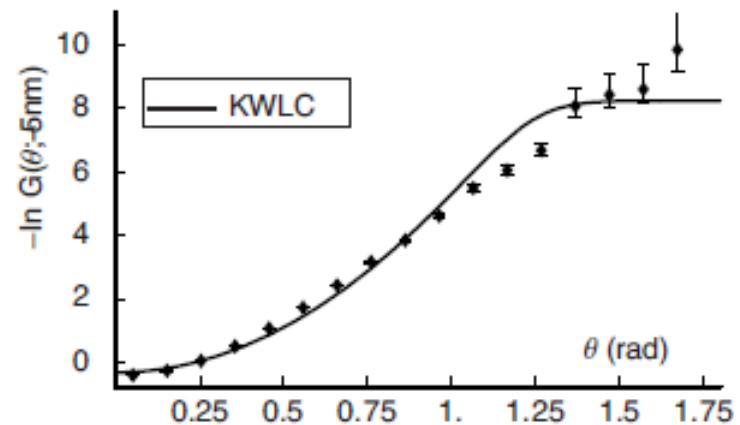
- Cyclization studies by Cloutier and Widom
- Sharply looped DNA highly bendable and also **highly twistable**, DNA more flexible than predicted by models
- Theories of DNA flexibility:

**Melted bubbles**, single-stranded regions with high flexibility, facilitate loop formation

## **Kinkable worm-like chain**

Extension of the WLC that includes spontaneous kink formation when curvature is high

⇒ Local alternative conformations with high flexibility



Wiggins, Nature Nanotechnology, 2006

# Development of new model

- Equation that fits measured data
- **Linear ‘sub-elastic chain’ (LSEC) model**

$$E_{\text{LSEC}}(\theta) = \alpha |\theta| k_B T$$

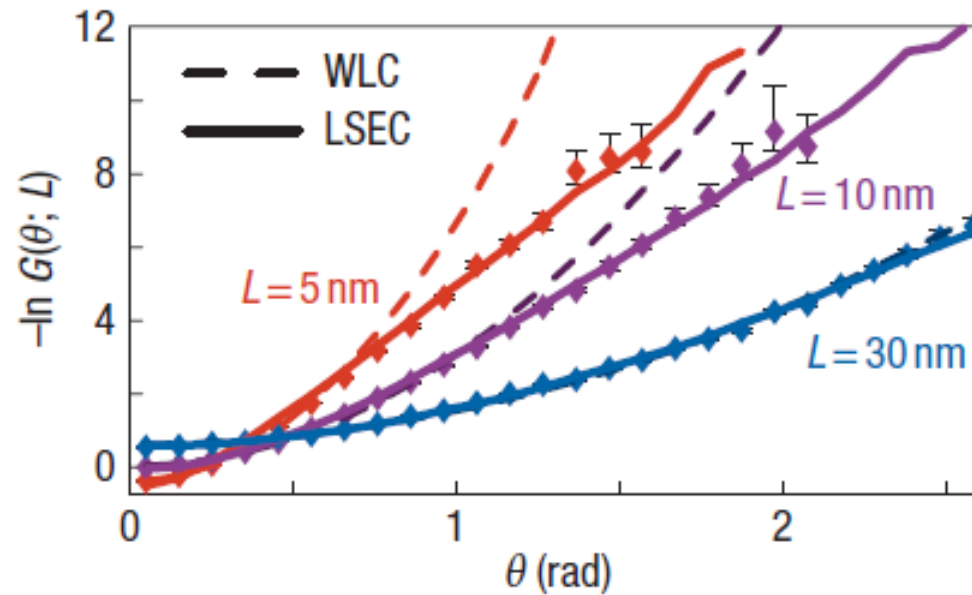
$\theta$ : bending angle between segments

$\alpha$ : dimensionless constant, dependent on  $\ell$

$k_B T$ : thermal energy

Non-harmonic (linear) energy function, “softer“ for sharp bending

# LSEC on long scale

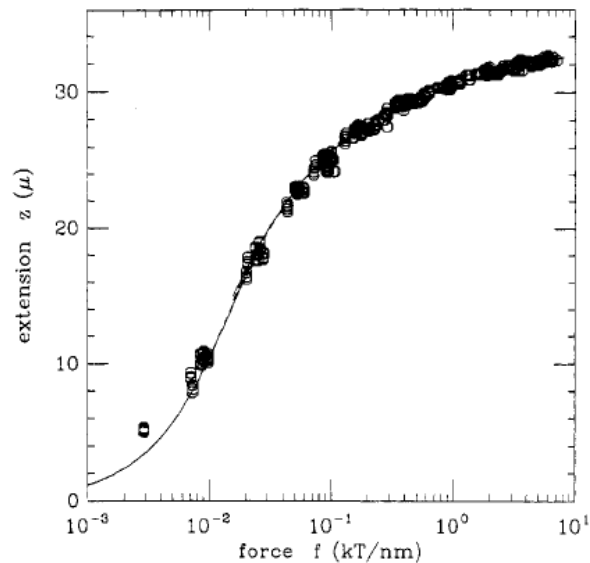


Wiggins, Nature Nanotechnology, 2006

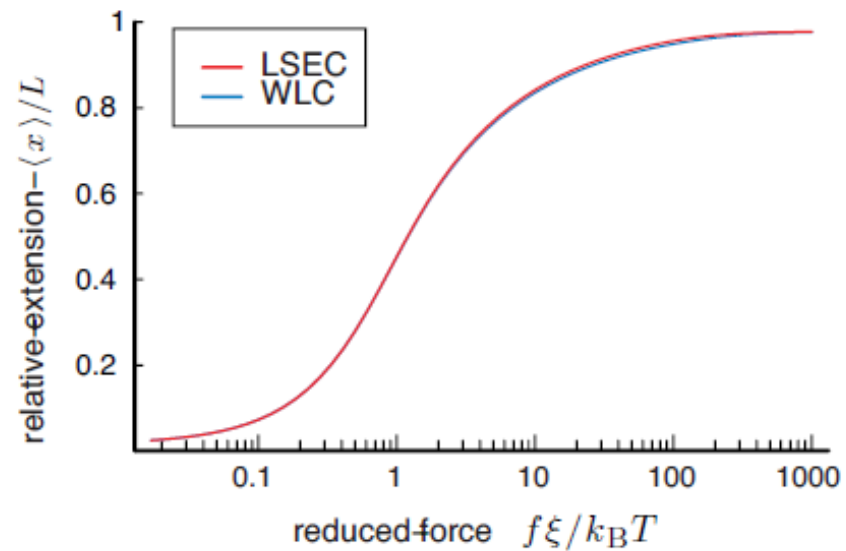
- WLC describes behaviour for **long** contour length correctly
- LSEC model fits data on **all** length scales

# LSEC on long scale

- Application of LSEC on other experimental data
- Force-extension experiment correctly described by LSEC



Marko and Siggia, *Macromolecules*, 1995



Wiggins, *Nature Nanotechnology*, 2006

# Outlook

## Improvement of new model

- Include twisting properties in LSEC model
- Test sequence dependence of DNA elasticity

## Application

- Relevance of model to other semiflexible biopolymers
  - Is DNA the only polymer with higher elasticity on short scales?

## Criticism

- Further/other experiments that support findings  
(Exclude that results are an artifact of the measurement)

# Summary

- DNA flexibility on short length scales is important in many biological processes (e.g. nucleosome formation).
- Using high-resolution AFM the bending behaviour of DNA in such scales can be measured directly.
- Sharp bends in DNA on short length scales are much more frequent than predicted by the WLC model.
- A simple model can describe both the bending behaviour of DNA on short scales and the long-length scale observations (force-extension curve of DNA)



# Literature

- Wiggins P.A., Van der Heijden T., Moreno-Herrero F., Spakowitz A., Phillips R., Widom J., Dekker C. and Nelson P.C. (2006) High flexibility of DNA on short length scales probed by atomic force microscopy. *Nature Nanotechnology* **1**, 137-141.
- Cloutier T.E. and Widom J. (2005) DNA twisting flexibility and the formation of sharply looped protein-DNA complexes. *Proc. Natl. Acad. Sci.* **102**,3645-3650.
- Podgornik R. (2006) Polymer Physics: DNA off the hook. *Nature Nanotechnology* **1**, 100-101.
- Wiggins P.A. and Nelson P.C. (2006) Generalized theory of semiflexible polymers. *Phys. Rev. E* **73**, 031906.
- Marko J.F. and Siggia E.D. (1995) Stretching DNA. *Macromolecules* **28**, 8759-8770.
- Swigon D., Coleman B.D., and Olson W.K. (2006) Modeling the Lac repressor-operator assembly: The influence of DNA looping on Lac repressor conformation. *Proc. Natl. Acad. Sci.* **103**, 9879–9884.
- Wiggins P.A., Philips R. and Nelson P.C.(2005) The exact theory of kinkable elastic polymers. *Phys. Rev. E* **71**, 021909.
- Yan J., Marko J.F. (2004) Localized single-stranded bubble mechanism for cyclization of short double helix DNA. *Phys. Rev. Lett.* **93**, 108108.
- Jena B.P. and Hörber J.K.H. (Editors) *Force microscopy: applications in biology and medicine* (J. Wiley & Sons, Hoboken, New Jersey, 2006)

Additional figures:

- <http://www.jyi.org/research/re.php?id=419>
- <http://www.nanomat.de/datenblaetter/42000.htm>
- <http://en.wikibooks.org/wiki/Nanotechnology/AFM>

**...Thank you for  
your attention!**